

09/581286

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MANAGER EXAMINATION SUPPORT AND

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AUSTRALIA

Patents Act 1990

CSL LIMITED

PROVISIONAL SPECIFICATION

Invention Title:

Porphyromonas gingivalis nucleotides

The invention is described in the following statement:

Porphyromonas gingivalis nucleotides

FIELD OF THE INVENTION

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The present invention relates to *P. gingivalis* nucleotide sequences, *P. gingivalis* polypeptides and probes for detection of *P. gingivalis*.

BACKGROUND OF THE INVENTION

Periodontal diseases are bacterial-associated inflammatory diseases of the supporting tissues of the teeth and range from the relatively mild form of gingivitis, the non-specific, reversible inflammation of gingival tissue to the more aggressive forms of periodontitis which are characterised by the destruction of the tooth's supporting structures. Periodontitis is associated with a subgingival infection of a consortium of specific Gram-negative bacteria that leads to the destruction of the periodontium and is a major public health problem. One bacterium that has attracted considerable interest is P. gingivalis as the recovery of this microorganism from adult periodontitis lesions can be up to 50% of the subgingival anaerobically cultivable flora, whereas P. gingivalis is rarely recovered, and then in low numbers, from healthy sites. A proportional increase in the level of P. gingivalis in subgingival plaque has been associated with an increased severity of periodontitis and eradication of the microorganism from the cultivable subgingival microbial population is accompanied by resolution of the disease. The progression of periodontitis lesions in non-human primates has been demonstrated with the subgingival implantation of P. gingivalis. These findings in both animals and humans suggest a major role for P. gingivalis in the development of adult periodontitis.

P. gingivalis is a black-pigmented, anaerobic, asaccharolytic, proteolytic Gram-negative rod that obtains energy from the metabolism of specific amino acids. The microorganism has an absolute growth requirement for iron, preferentially in the form of haeme or its Fe(III) oxidation product haemin and when grown under conditions of excess haemin is highly virulent in experimental animals. A number of virulence factors have been implicated in the pathogenicity of P. gingivalis including the capsule, adhesins, cytotoxins and extracellular hydrolytic enzymes. In

particular, proteases have received a great deal of attention for their ability to degrade a broad range of host proteins including structural proteins and others involved in defence. The proteins that have been shown to be substrates for P. gingivalis proteolytic activity include collagen types I and IV, fibronectin, fibrinogen, laminin, complement and plasma clotting cascade proteins, α_1 -antitrypsin, α_2 -macroglobulin, antichymotrypsin, antithrombin III, antiplasmin, cystatin C, IgG and IgA. The major proteolytic activities associated with this organism have been defined by substrate specificity and are "trypsin-like", that is cleavage on the carboxyl side of arginyl and lysyl residues and collagenolytic although other minor activities have been reported.

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P. gingivalis trypsin-like proteolytic activity has been shown to degrade complement, generating biologically active C5a, impair the phagocytic and other functions of neutrophils by modifying surface receptors, and abrogate the clotting potential of fibrinogen prolonging plasma clotting time. The trypsin-like proteolytic activity of P. gingivalis also generates Fc fragments from human IgG1 stimulating the release of proinflammatory cytokines from mononuclear cells and is associated with vascular disruption and enhanced vascular permeation through the activation of the kallikrein-kinin cascade. P. gingivalis spontaneous mutants with reduced trypsin-like activity as well as wild-type cells treated with the trypsin-like protease inhibitor N-p-tosyl-L-lysine chloromethyl ketone are avirulent in animal models. Further, it has been shown that P. gingivalis grown under controlled, haemin-excess conditions expressed more trypsinlike and less collagenolytic activity and were more virulent in mice relative to cells grown under haemin-limited but otherwise identical conditions. The increased expression of the trypsin-like activity by the more virulent P. gingivalis has led to the speculation that the trypsin-like proteolytic activity may be the major determinant for infection or disease.

There has been considerable endeavour to purify and characterise the trypsin-like proteases of *P. gingivalis* from cell-free culture fluids. Chen et al, (1992) [J Biol Chem 267:18896-18901] have purified and characterised a 50 kDa arginine-specific, thiol protease from the culture fluid of *P. gingivalis* H66 designated Arg-gingipain. A similar arginine-specific thiol protease has been disclosed in JP 07135973 and the amino acid sequence disclosed in WO 9507286 and in Kirszbaum et al, 1995 [Biochem Biophys Res Comm

207:424-431]. Pike et al (1994) [J Biol Chem 269:406-411] have characterised a 60 kDa lysine-specific cysteine proteinase from the culture fluid of P. gingivalis H66 designated Lys-gingipain and the partial gene sequence for this enzyme was disclosed in WO 9511298 and fully disclosed in WO 9617936.

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In order to develop an efficacious and safe vaccine to prevent P. gingivalis colonisation it is necessary to identify and produce antigens that are involved in virulence that have utility as immunogens to generate neutralising antibodies. Whilst it is possible to attempt to isolate antigens directly from cultures of P. gingivalis this is often difficult. For example as mentioned above, P. gingivalis is a strict anaerobe and can be difficult to isolate and grow. It is also known that, for a number of organisms, when cultured in vitro that many virulence genes are down regulated and the encoded proteins are no longer expressed. If conventional chemistry techniques were applied to purify vaccine candidates potentially important (protective) molecules may not be identified. With DNA sequencing, as the gene is present (but not transcribed) even when the organism is grown in vitro it can be identified, cloned and produced as a recombinant DNA protein. Similarly, a protective antigen or therapeutic target may be transiently expressed by the organism in vitro or produced in low levels making the identification of these molecules extremely difficult by conventional methods.

With serological identification of therapeutic targets one is limited to those responses which are detectable using standard methods such as Western Blotting or ELISA. The limitation here is the both the level of response that is generated by the animal or human and determining whether this response is protective, damaging or irrelevant. No such limitation is present with a sequencing approach to the identification of potential therapeutic or prophylactic targets.

It is also well known that *P. gingivalis* produces a range of broadly active proteases (University of Melbourne International Patent Application No PCT /AU 96/00673, US Patent Nos 5,475,097 and 5,523,390), which make the identification of intact proteins difficult because of their degradation by these proteases.

SUMMARY OF THE INVENTION

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The present inventors have attempted to isolate P. gingivalis nucleotide sequences which can be used for recombinant production of P. gingivalis polypeptides and to develop nucleotide probes specific for 5 P. gingivalis. The DNA sequences listed below have been selected from a large number of P. gingivalis sequences according to their indicative potential as vaccine candidates. This intuitive step involved comparison of the deduced protein sequence from the P. gingivalis DNA sequences to the known protein sequence databases. Some of the characteristics used to 10 select useful vaccine candidates include; the expected cellular location, such as outer membrane proteins or secreted proteins, particular functional activities of similar proteins such as those with an enzymatic or proteolytic activity, proteins involved in essential metabolic pathways that when inactivated or blocked may be deleterious or lethal to the organism, proteins -15 that might be expected to play a role in the pathogenesis of the organism eg. red cell lysis, cell agglutination or cell receptors and proteins which are paralogues to proteins with proven vaccine efficacy. DNA sequences that were considered to be poor vaccine candidates and not selected include those that code for proteins involved in replication, non-essential proteins 20 involved in cellular processes and those proteins present at sites that would be unlikely to be affected by immune mediators such as those found in the bacterial cytoplasm or inner membranes.

In a first aspect the present invention consists in an isolated *P. gingivalis* nucleotide sequence, the nucleotide sequence consisting of or including a sequence selected from the group consisting of SEQ ID NO: 1, SEQ ID NO: 2, SEQ ID NO: 3, SEQ ID NO: 4, SEQ ID NO: 5, SEQ ID NO: 6, SEQ ID NO: 7, SEQ ID NO: 9, SEQ ID NO: 10, SEQ ID NO: 11, SEQ ID NO: 12, SEQ ID NO: 13, SEQ ID NO: 14, SEQ ID NO: 15, SEQ ID NO: 16, SEQ ID NO: 17, SEQ ID NO: 18, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 21, SEQ ID NO: 22, SEQ ID NO: 23, SEQ ID NO: 24, SEQ ID NO: 25, SEQ ID NO: 26, SEQ ID NO: 27, SEQ ID NO: 28, SEQ ID NO: 29, and SEQ ID NO: 30.

In a second aspect the present invention consists in an isolated *P. gingivalis* polypeptide, the polypeptide being at least partially encoded by a nucleotide consisting of or including a sequence selected from the group

consisting of SEQ ID NO: 1, SEQ ID NO: 2, SEQ ID NO: 3, SEQ ID NO: 4, SEQ ID NO: 5, SEQ ID NO: 6, SEQ ID NO: 7, SEQ ID NO: 9, SEQ ID NO: 10, SEQ ID NO: 11, SEQ ID NO: 12, SEQ ID NO: 13, SEQ ID NO: 14, SEQ ID NO: 15, SEQ ID NO: 16, SEQ ID NO: 17, SEQ ID NO: 18, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 21, SEQ ID NO: 22, SEQ ID NO: 23, SEQ ID NO: 24, SEQ ID NO: 25, SEQ ID NO: 26, SEQ ID NO: 27, SEQ ID NO: 28, SEQ ID NO: 29, and SEQ ID NO: 30.

In a third aspect the present invention consists in a nucleotide probe specific for *P. gingivalis*, the probe including a detectable label and a nucleotide sequence of at least 15(?) nucleotides, the nucleotide sequence being derived from a sequence selected from the group consisting of SEQ ID NO: 1, SEQ ID NO: 2, SEQ ID NO: 3, SEQ ID NO: 4, SEQ ID NO: 5, SEQ ID NO: 6, SEQ ID NO: 7, SEQ ID NO: 9, SEQ ID NO: 10, SEQ ID NO: 11, SEQ ID NO: 12, SEQ ID NO: 13, SEQ ID NO: 14, SEQ ID NO: 15, SEQ ID NO: 16, SEQ ID NO: 17, SEQ ID NO: 18, SEQ ID NO: 19, SEQ ID NO: 20, SEQ ID NO: 21, SEQ ID NO: 22, SEQ ID NO: 23, SEQ ID NO: 24, SEQ ID NO: 25, SEQ ID NO: 26, SEQ ID NO: 27, SEQ ID NO: 28, SEQ ID NO: 29, and SEQ ID NO: 30, or a sequence complementary thereto.

20 DETAILED DESCRIPTION

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Preparation of the P. gingivalis library for sequencing.

To determine the DNA sequence of P. gingivalis genomic DNA was isolated 25 from P. gingivalis strain W50 (ATCC 53978) essentially by the method described by Mamur J. (1961). Cloning of DNA fragments was performed essentially as described by Fleischmann et al., (1995). Briefly, purified genomic DNA from P. gingivalis was nebulized to fragment the DNA and was treated with Bal31 nuclease to create blunt ends then run twice on 30 preparative 1% agarose gels. DNA fragments of 1.6-2.0 kb were excised from the gel and the DNA recovered. This DNA was then ligated to the vector pUC18 (Smal digested and dephosphorylated; Pharmacia) and electrophoresed on a 1% agarose preparative gel. The fragment comprising linear vector plus one insert was excised, purified and this process repeated 35 to reduce any vector without insert contamination. The recovered vector plus insert DNA was blunt-ended with T4 DNA polymerase, then a final

ligation to produce circular DNA was performed. Aliquots of Epicurian Coli Electroporation-Competent Cells (Stratagene) were transformed with the library DNA and plated out on SOB agar antibiotic diffusion plates containing X-gal and incubated at 37°C overnight. Colonies with inserts appeared white and those without inserts (vector alone) appeared blue. Plates were stored at 4°C until the white clones were picked and expanded for the extraction of plasmid DNA for sequencing.

DNA sequencing

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Plasmid DNA was prepared by picking bacterial colonies into 1.5ml of LB, TB or SOB broth supplemented with 50-100ug/ml Ampicillin in 96 deep well plates. Plasmid DNA was isolated using the QIAprep Spin or QIAprep 96 Turbo miniprep kits (QIAGEN GmbH, Germany). DNA was eluted into a 96 well gridded array and stored at -20C.

Sequencing reactions were performed using ABI PRISM Dye Terminator and ABI PRISM BIGDye Terminator Cycle Sequencing Ready Reaction kits with AmpliTaq DNA polymerase FS (PE Applied Biosystems, Foster City, CA) using the M13 Universal forward and reverse sequencing primers. Sequence reactions were conducted on either a Perkin-Elmer GeneAmp 9700 (PE Applied Biosystems) or Hybaid PCR Express (Hybaid, UK) thermal cyclers. Sequencing reactions were analysed on ABI PRISM 377 DNA sequencers (PE Applied Biosystems).

The sequences obtained are set out below.

25 DNA sequence analysis

Raw trace data files from the ABI 377 sequencer were manually trimmed using Staden Pregap(Laboratory of Molecular Biology, Medical Research Council, UK) running on a Sun Microsystem computer. Trimmed files were assembled into contigs using Staden Gap v4.1 and exported as a consensus file in FastA format. This consensus was converted into GCG format files and analysed for homology using the BLASTX algorithm [Altschul et al] on a non-redundant protein database compiled by ANGIS (Australian Genomic Information Service, University of Sydney). Individual BLAST search results were examined for significant homology by statistical probability and amino acid alignments.

The results are set out in Table 1.

It will be appreciated by persons skilled in the art that numerous variations and/or modifications may be made to the invention as shown in the specific embodiments without departing from the spirit or scope of the invention as broadly described. The present embodiments are, therefore, to be considered in all respects as illustrative and not restrictive.

Dated this tenth day of December 1997

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CSL LIMITED

Patent Attorneys for the Applicant:
F.B. RICE & CO.

References.

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Mamur, J. (1961) A procedure for the isolation of deoxyribonucleic acid from micro-organisms. J. Mol. Biol. 3, 208-218.

Fleishmann, R.D. et al. (1995) Whole genome random sequencing and assembly of *Haemophilus influenzae* Rd. Science 269, 496-512.

Altschul SF, Gish W, Miller W and EW Myers. (1990). Basic local alignment search tool. J. Mol. Biol. 215:403-410.

SEQUENCES

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Seq ID # 15
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     ctcatccaca acgatggaga catgcacctt attggctctg aactcctcga
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     gcaaatcatc tatgcgcttg ttttcgggga caaaatatgc tttacgaatc
251
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     tttgatgtaa atcaccctt tgatattgtc ttctgacccc tctgaaacgg
351 gaagtetgga ataaceegae gaaacaaega agteaageat ettaegaaat
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451 tcgcaggctg gcttattata ggaattg
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Length: 486
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 201 accatggate tecacagaag egaeggeagt agggteaceg gatgtagtea
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 451 aatggcgaac tcctttggta atcatacccg ggaaat
Seq ID # 17
Length: 386
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  51 gccgtagcga tggagggcat tcgcccgata ctcatcgaag cgcangcttt
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151 atattcggcg gatgaacatg ctcttagccg tactggagaa acgtgccggc
201 ttcaagctca tacagaanga tgtgtttctg aacattgccg gaggtatcaa
251
     aatagccgat ccggctacgg atctggccgt tatctcggca gtgctggcgt
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     ccagtctgga catcgttatc ccgccggccg tatgcatgac gggcgaagtc
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Seq ID # 18 Length: 1013

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      ggatgaggct ttggagaaat tggctcgtct gggttataag aagatcaatg
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Seq ID # 20
Length: 488
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      tcagtcggat atagtcatcc gattcgtagt ccgtacgtcc ggccatttca
 151
      toggacagac googtatoto ttoototatt tggcgaatat cgttgaaagc
      ctgctcgacc tcttcgtaaa ccgtgtgtcc gtcctgcaaa cgcatcacct
 201
 251
      gcggcagata gcctatgcgg atccccttgg ggcgtgctat gtgtccggat
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      acceptante cotacaagag cgatacggtc gcgcctgttg atgacgaatg
 351
 401
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Seq ID # 21
Length: 836
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 151 ccggctattc cgagactcct taaggagtcc ctaccgagac ccttaaggag
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      ggaagcette acacceagat ngacgagegg caactecaeg ttetegetta
701
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Seq ID # 22
Length: 365
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     caatggaatc gacctgctcg aactcgaacc ggaagaacgt gcacacctcg
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351

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 251
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401

451

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Seq ID # 28 Length: 740

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551

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Seq ID#	Description	Accession	%	overlap
-	48kD outer membrane protein of Actinobacillus pleuropneumoniae	number	identity	(aa)
7	Eukaryote outer membrane protein. TCR iunction seguence	E250352	F. C. Z	071
m	Perinjasmic zinc metallomentease belomeing to the inculiance femilians	2227332	40.7	113
, -	sapiration and an appropriate and an arrangement of the modification of the contract of the co	P37648	22.2	126
4	Heat-shock protein DNAJ of Legionella pneumophila	P50025	42.2	173
٧	Eukaryote plasma cell membrane glycoprotein (alkaline phosphodiesterase I)	P22413	42.6	101
9	Alpha-hemolysin gene, hlyA, Aeromonas hydrophila	L36462	55.6	36
7	Protein translocase SecA subunit, Escherichia coli	P10408, P75642	40.7	3 ==
∞	Protein-export membrane protein (secD), Helicobacter pylori	AE000652 12	41.2	¥.
6	Hypothetical integral membrane protien, Helicobacter pylori	AE000647 20	29.2	99
10	ATP-dependent CLPC protease, Mycobacterium leprae	_ P24428	40.1	17
=	Zinc-carboxypeptidase precursor, Streptomyces capreolus	P39041	30.3	66
12	Hem B receptor, Porphyromonas gingivalis	U87395	37.6	109
13	Protein export protein, Helicobacter pylori	AE000652 11	45.8	96
14	Haemoglobin receptor, Neisseria gonorrhoeae	F72073	28.7	115
15	Haemolysin, Helicobacter pylori	AE000647 24	37.7	146
91	Outer membrane protein, Bacteroides thetaiotaomicron	Q45780	39.9	168
11	ATP-dependent protease, Helicobacter pylori	AE000542 5	40.7	123
<u>∞</u>	Acid phophatase precursor, Flavobacterium meningosepticum	008351	41.1	129
19	CBIK protein involved in Cobalmin biosynthesis, Salmonella typhimurium	005592	42.3	64
20	ABC Transporter, Haemophilus influenzae	005519	34	153
21	ABC transporter, Bacillus subtlis	AF008220 56	49.3	240
22	ABC transporter, Mycobacterium leprae	E343546	56.3	87
23	Cysteine protease, Trypanosoma brucei	012000	, , ,	

Table 1 (cont.)

Seq ID#	Description	Accession	%	overlap
24	Suface antigen gene, Methanosarcina mareii	number X84710	identity 42.9	(aa) 126
25	Protein-export membrane protein SECD, Haemophilus influenzae	P44591	51.5	132
26	Hypothetical protein involved in Cobalamin synthesis, Methanococcus jannaschii	Q60342	37.5	168
27	Haem uptake protein B, Bacteroides fragilis	Q45140	53	99
. 28	Virulence-associated ABC transporter, Francisella novicida	Q47909	36.4	66
29	Hypothetical secreted protein, Helicobacter pylori	AE000535_9	28.4	102
30	ABC transporter FTSE, Escherichia coli	P10115	48.4	64